PII: S0040-4039(96)01806-0

## **Inter-Flavin Electron Transfer in the Ground State** in the Absence of External Electron Donors and Acceptors

Humphrey I. X. Mager<sup>1</sup> and Shiao-Chun Tu<sup>1,2</sup>

Departments of <sup>1</sup>Biochemical and Biophysical Sciences and <sup>2</sup>Chemistry, University of Houston, Houston, TX 77204-5934, U.S.A.

Abstract: Inter-flavin electron transfer of N5-alkylflavinium cation 3 occurred in the ground state to primarily give the flavosemiquinone 4 and the superoxidized flavin 7. This provided insight into some chemistry of 7: a rapid addition followed by a (second) inter-flavin electron transfer of the derived  $4^a$ -flavin adduct radical 6 giving 3 and, consequently, an additional formation of 4. Copyright © 1996 Elsevier Science Ltd

In the flash photolysis of lumiflavin, excitation of a preformed dimer or more complex aggregate was formulated 1 to proceed through the singlet and triplet states resulting in a splitting of the dimer or aggregate to the flavosemiquinone anion and a flavin radical cation (Eq. 1). Formation of a flavin radical cation in aqueous solution can take place via photoionization. 2 The reaction of the flavin triplet states with another flavin molecule either in the triplet or ground state has been concluded 3 to occur via inter-flavin electron transfer to give the Fl<sup>-\*</sup> and Fl<sup>+\*</sup> radicals (Eq. 2). In

$$2Fl = Fl_2 \xrightarrow{hv} {}^{1}Fl_2 \longrightarrow {}^{3}Fl_2 \longrightarrow F\overline{l} + F\overline{l} + F\overline{l} + (1)$$

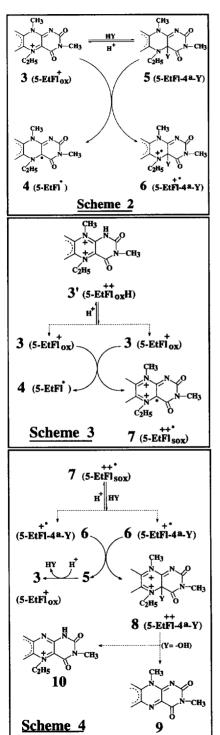
$${}^{3}Fl + Fl \longrightarrow F\overline{l} + F\overline{l} + (2)$$

$$Fl_{0x} = Fl_{sox} + e^{-} \qquad (3)$$

$${}^{R}_{10x} \longrightarrow {}^{R}_{10x} \longrightarrow {$$

the absence of an external electron donor,  $FI^{++}$  rapidly reacts with  $H_2O$ , but the product(s) so formed was (were) not identified. The authors<sup>3</sup> did not spectrophotometrically detect  $FI^{++}$ , but Heelis and coworkers<sup>4</sup> later succeeded in determining the spectral characteristics of this radical which, in contrast with the earlier assumption,<sup>3</sup> did absorb significantly at  $\lambda > 550$  nm. Regarding the oxidation state of the  $FI^{++}$  radical, the term "superoxidized" flavin (abbreviated by the subscript "sox") has been introduced<sup>5</sup> to emphasize the existence of an oxidation state higher than the one of the so-called normal "fully oxidized" flavin (abbreviated as  $FI_{ox}$ ; Eq. 3). The superoxidized flavin 1 and derived 4<sup>a</sup>-hydroxy-4<sup>a</sup>,5-dihydroflavin radical cation 2 are assumed to be key-intermediates in bacterial bioluminescence.<sup>5-7</sup> The existence of flavin derivatives related to 1 and 2 (cf. 7 and 6 [X= OH; OMe]) was proven by applying cyclic voltammetry,<sup>8</sup> while 6 or a derivative has appeared to accomplish the chemical hydroxylation of aromatic substrates.<sup>9</sup>

The present studies have shown that inter-flavin electron transfer comparable with the one represented by Eq. 2 may occur in the ground state when started from N5-alkylflavinium cations (e.g. 3; Scheme 3).

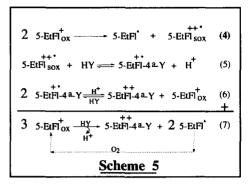


Quantitative studies on the formation of the 5-ethylflavosemiquinone 4 have substantiated some chemistry of the superoxidized flavin 7: a rapid addition  $(7 + HY \rightarrow 6 + H^+; Y^- = HO^-, Cl^-, Br^-; Scheme 4)$  followed by a (second) inter-flavin electron transfer of the derived  $4^a$ -flavin adduct radical 6 giving 3 and, consequently, an additional generation of 4.

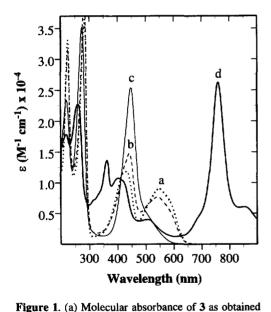
In previous studies on the spontaneous formation of the flavosemiquinone 4 (Scheme 2), the flavinium cation 3 has been considered to be the acceptor of an electron  $(3 + e^{-} + 4)$  donated by a  $4^a$ -flavin adduct  $5 (\rightarrow 6 + e^{-})$  both in organic and aqueous solutions. <sup>10</sup> The flavinium cation 3 is in equilibrium with the dihydroflavin pseudobase  $5 (Y = OH; pK'_a = 4.15 \text{ in } 0.1 \text{ M}$  citrate buffer). In aqueous solution, in the pH 2 - 7 range at 23 °C a relative initial rate / pH profile was found <sup>10d</sup> to be in agreement with the view depicted in Scheme 2. Consistently, the rate of the radical formation, as indicated by the appearance of 4, decreased upon increasing the pH in the pH 4.1 - 7.0 range and upon lowering the pH in the pH 4.1 - 2.0 range.

However, further increase of the acidity up to 12 N, using H<sub>2</sub>SO<sub>4</sub>, HClO<sub>4</sub>, HCl and HBr, caused a remarkable increase of the rate of the radical formation indicating the occurrence of a different pathway in which the flavin pseudobase was no longer involved. Although slower, spontaneous radical formation even occurred when 3 was shifted to the protonated species 3' (Scheme 3) as indicated by the appearance of an alloxaziniumlike absorption spectrum (Fig. 1; curve a → b → c) and by increased down-field shifts of the signals for the N<sup>10</sup>-CH<sub>3</sub> and aromatic protons in the PMR-spectra [3 (CD<sub>3</sub>CN): δ= 1.93 (3, t, J= 8 Hz, C-Me); 2.57 (3, s, C-Me); 2.63 (3, s, C-Me); 3.41 (3, s, N-Me); 4.17 (3, s, N<sup>10</sup>-Me); 5.00 (1, C-H); 6.05 (1, C-H); 7.94 (1, s, Ar-H); 8.20 (1, s, Ar-H). For 3' (CD<sub>3</sub>CN / 5%  $H_2SO_4$ ):  $\delta = 1.83$  (3, t, C-Me); 2.71 (3, s, C-Me); 2.76 (3, s, C-Me) Me); 3.51 (3, s, N-Me); 4.52 (3, s, N<sup>10</sup>-Me); 8.34 (1, s, Ar-H); 8.53 (1, s, Ar-H)]. Spontaneous formation of flavin radicals also took place in acidified organic solutions of 3. Under some conditions, the reaction times could even be reduced to less than 30 minutes at room temperature (Fig. 2). The flavosemiquinone 4 is stable in the absence of O<sub>2</sub> thus allowing accurate analyses of it based on molecular extinctions established. 10b The yields of 4, formed anaerobically under variable experimental conditions in the absence of any additional compound were found to be in the range of 64 - 69% (cf. Fig. 2, curve f). These results are in good agreement with the sequence of reactions formulated to proceed as summarized in Scheme 5 (Eqs. 4 - 6):

(Eq. 4): A primary inter-flavin electron transfer of the flavinium cation 3 (Scheme 3) to give the flavosemiquinone 4 and, consequently, the superoxidized flavin 7. (Eq.5): The addition of H<sub>2</sub>O to the superoxidized flavin 7 leading to the formation of the radical cation 6 (Y= OH;



Scheme 4). (Eq. 6): An inter-flavin electron transfer of 6 (Scheme 4) resulting in the flavinium cation 3 and a bicationic product 8 which was already observed in the electrochemical oxidation of 5 (Y= OH; OMe) by the release of 2 x 1 electron in two distinct steps. (Eq. 7): As the important consequence of Scheme 5, the overall anaerobic transformation of the flavinium cation 3, in the absence of other compounds, implies the formation of the flavosemiquinone 4 in a calculated, maximal yield of 66.7 % and of the bicationic product 8 in a calculated maximal yield of 33.3 %.



from a 5 x 10<sup>-5</sup> M solution in MeCN,  $\lambda_{\text{max}}$  at 224, 283, 432 and 548 nm. (b) Spectrum of a mixture of species 3 and 3' in 12 N H<sub>2</sub>SO<sub>4</sub>. (c) Molecular absorbance of 3' in 11.6 N HClO<sub>4</sub> (complete protonation),  $\lambda_{\text{max}}$  at 219, 275 and 448 nm. (d) Spectrum of the new chloride in 12 N H<sub>2</sub>SO<sub>4</sub> with  $\lambda_{\text{max}}$  at 363, 397, 511, 761 and 853 nm used as criteria for the purity of the compound.

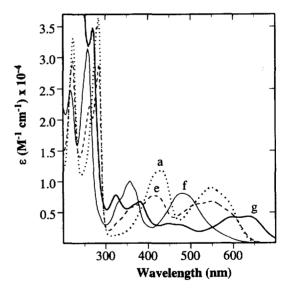


Figure 2. (a) See Figure 1; (e) Spontaneous change of spectrum at 23 °C, 3 min after the addition of conc HCl (HCl/flavin ratio=10). (f) Spectrum of a mixture of protonated flavosemiquinone 4 and product(s) derived from 8 at the end of the spontaneous, anaerobic process, 30 min after the addition of HCl. (g) Spectral change following the addition of NEt<sub>3</sub> (0.1 vol %) to convert the protonated flavosemiquinone to non-protonated 4 revealing its accurate yield (65.5%).

The electron-deficient nature of 8, in these experiments formally derived from the flavin adduct by the loss of two electrons seems to be supported by the structures of the products of a ready decomposition leading to 3-methyllumiflavin 9 and 5-ethyl-3-methylalloxazinium cation 10 (Scheme 4). In order to increase the yield of 8, 5-EtFl was recycled to 5-EtFl by air-oxidation (Eq. 7). When both species 4 and 3 were no longer detected, the solution of 8 or a derived product was still found able to achieve the hydroxylation of an aromatic substrate. 9

In comparable acidic environment, the presence of Cl $^{-}$  or Br $^{-}$  caused a considerable increase of the radical formation rate. We suggest that this enhancing anion effect is due to a faster addition of Cl $^{-}$  (Br $^{-}$ ) to provide different radical cations 6 (Eq. 5; Y= Cl, Br). In the presence of H<sub>2</sub>O, an exchange of Cl $^{-}$  (Br $^{-}$ ) by HO $^{-}$  may readily occur in a further stage of the process.

From a more concentrated solution (≈ 10<sup>-1</sup> M) of 3 in MeCN / HCl, a new solid chloride was isolated which rapidly decomposed in alcoholic and aqueous solutions to give 3-methyllumiflavin 9 as a major product. The decomposition slowed down in 12 N H<sub>2</sub>SO<sub>4</sub> to give a remarkable absorption spectrum with high absorbances at higher wavelengths (Fig. 1, curve d). The new compound has reversible redox properties. Tentative mass spectral studies showed the presence of a peak at m/e 334 suggesting the covalent binding of a chlorine atom by the flavin. This peak probably represents a fragmentation of a dimer of a yet unknown oxidation level, in the sense that no conclusion is yet to be drawn whether the product has been derived from 6 or 8. It is emphasized that a similar spectrum can be obtained in the absence of Cl indicating the existence of more than one product with a comparable structure. Dimerization implies the relative stabilization of an intermediate which is also of interest in connection with the early view that interflavin electron transfer would take place in a dimer (Eq. 1) or more complex aggregate. The structure elucidation of the new chloride is in progress.

Acknowledgements. This work was supported by grants GM 25953 from NIH and E-1030 from the Robert A. Welch Foundation.

## References

- 1. Ballard, S. G.; Mauzerall, D. C.; Tollin, G. J. Phys. Chem. 1976, 80, 341 351.
- 2. Getoff, N.; Solar, S.; McCormick, D. B. Science 1978, 201, 616 618.
- 3 Hemmerich, P.; Knappe, W.R.; Kramer, H. E. A.; Traber, R. Eur. J. Biochem. 1980, 104, 511 520.
- 4. Heelis, P. F.; Parsons, B. J.; Phillips, G. O.; Swallow, A. J. J. Phys. Chem. 1986, 90, 6833 6836.
- 5. Mager, H. I. X.; Tu, S.-C.; Liu, Y.-H.; Deng, Y.; Kadish, K. M. *Photochem. and Photobiol.* 1990, 52, 1049 1056.
- 6. Merényi, G.; Lind, J. L.; Mager, H. I. X.; Tu, S.-C. J. Phys. Chem. 1992, 96, 10528-10533.
- 7. Mager, H. I. X.; Tu, S.-C, Photochem. and Photobiol. 1995, 62, 607 614; and reference cited therein.
- 8. Mager, H. I. X.; Sazou, D.; Liu, Y.-H.; Tu, S.-C.; Kadish, K.M. J. Am. Chem. Soc. 1988, 110, 3759 -3762.
- 9. (a) Mager, H. I. X.; Tu, S.-C, Tetrahedron 1994, 50, 5287 5298; (b) Tetrahedron 1994, 50, 6759 6766.
- (a) Mager, H. I. X.; Addink. R. Flavins and Flavoproteins (Bray, R. C.; Engel, P.C.; Mayhew, S. G., Eds., Walter de Gruyter, Berlin) 1984, 37 40; (b) Tetrahedron 1985, 41, 183 190; (c) Mager, H. I. X.; Tu, S.-C, Flavins and Flavoproteins (Edmondson, D. E.; McCormick. D. B., Eds., Walter de Gruyter, Berlin) 1987, 583 592; (d) Tetrahedron 1988, 44, 5669 5674.